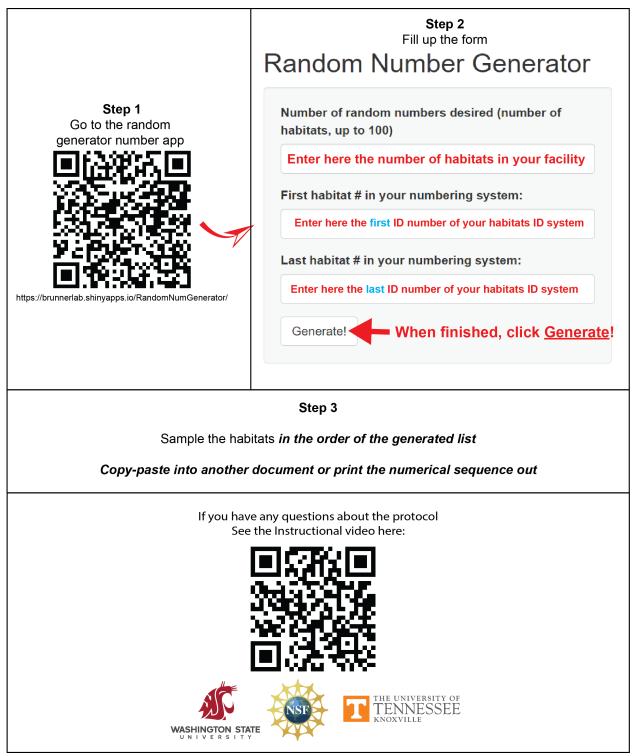


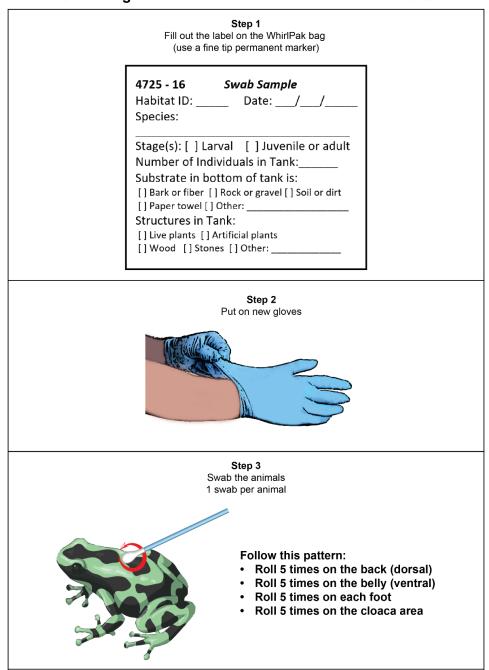
Pet Amphibian Trade Pathogen Testing Study Instructions for sampling

1. Randomizing/selecting habitats to sample

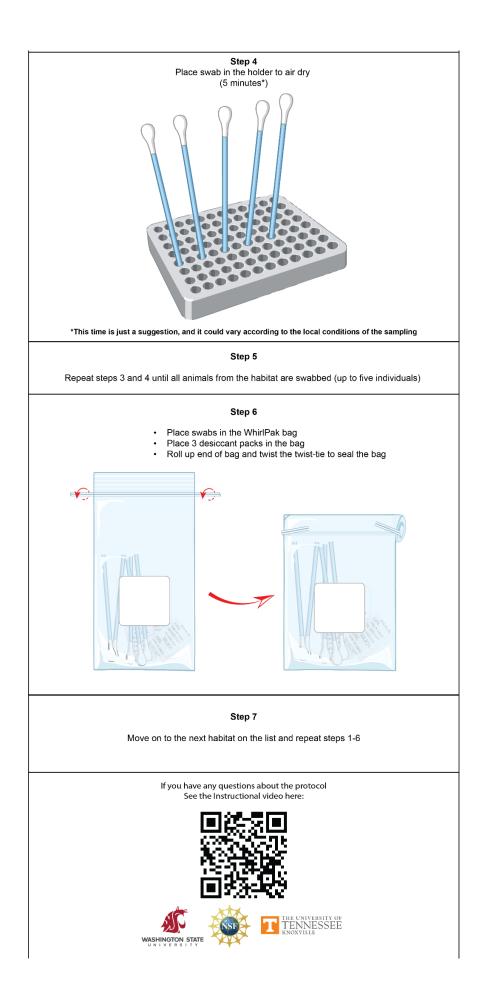




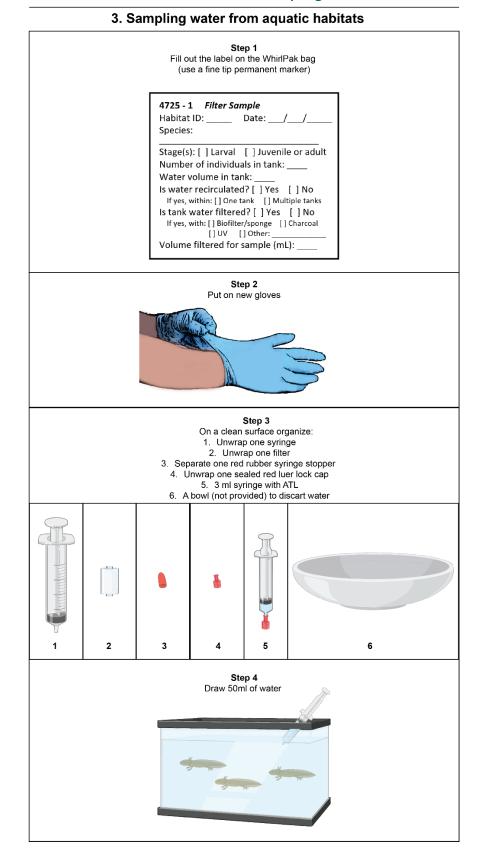
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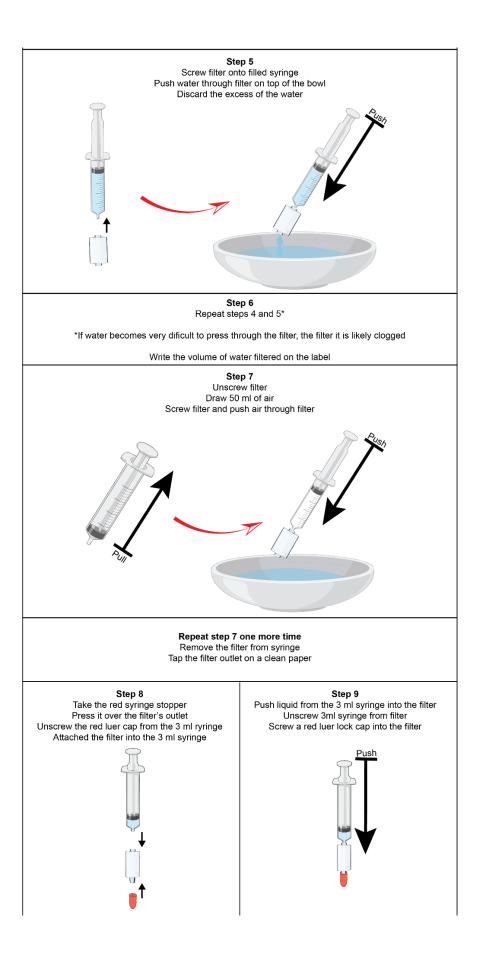


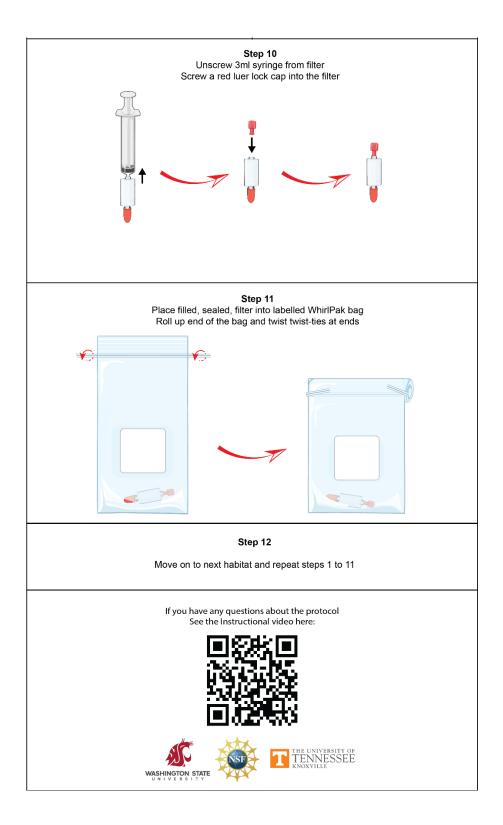
2. Swabbing individual animals from terrestrial habitats



Pet Amphibian Trade Pathogen Testing Study Instructions for sampling







Preparing for shipping

- Create and write down a unique PIN (4-8 digits is fine) on the included sheet
 - Keep one copy and send the other to us
 - This is the only way to retrieve your results, so please be sure not to lose the PIN
- Ensure all bags are closed and *labelled*
- Place all bagged samples in padded return envelope
- Make sure return label is firmly affixed
- Place in any outgoing mail drop

Questions?

• You can ask questions or obtain additional supplies by contacting Ashley Brinkman at Ashley@petadvocacy.org or (202) 452-1525 extension 1010

Thank you!

Pet Advocacy Network Pet Amphibian Trade Pathogen Testing Study Detailed instructions for sampling

Randomizing/selecting habitats to sample

Instructional video here:



https://youtu.be/CeCwUUq7xm4

Rational: We want to get an unbiased, representative sample from your facility. We therefore need to make sure that a random selection of habitats are sampled. Habitats are defined as tanks, containers, or groups of housing containers between which water or animals can move.

- 1. Go to: https://brunnerlab.shinyapps.io/RandomNumGenerator/
- 2. Enter the number of habitats in your facility (and optionally, the starting and ending number, if you already have a numbering scheme)
 - It does not matter how you number your habitats, just so that they are numbered in some sequential way
- 3. Click "Generate!", and then print out the list
- 4. Sample the tanks in the order of this list
 - You will likely not get to the end of the list with the materials we have provided. That is OK. Just go until you run out of materials or you can't stand this exercise any more... We'll take whatever you can give
 - You may run out of swabs before you run out of filters, or vice versa. This is OK, too. Just move on to the next tank in the list and sample it if you can

Swabbing individual animals from terrestrial habitats

Instructional video here:



https://youtu.be/cye9x4a-zJs

Overview: We ask you to swab the skin of *up to* five individuals per habitat. If there is one animal, swab that one. If there are 30, sample the first five you catch. If you run out of supplies, sample as many as you can.

- 1. Wear new gloves for each habitat. You do not need to change gloves between individuals within a habitat
 - If you have sweaty or clammy hands, you may wish to double-glove to make it easier to change the outer pair of gloves between tanks
- 2. Fill out the label on the WhirlPak bag (please use a fine tip permanent marker as pens will smudge) (Figure 1)
 - Habitat ID (to relate results back to a habitat)
 - Species (Latin binomial, please)
 - Life stage(s) (Larva vs. adult)
 - Number of individuals in the tank
 - Approximate surface area of tank bottom
 - Please provide units! (Metric is preferred, but we can convert)
 - Type of substrate in the tank (bark or fiber, soil or dirt, paper towel, or other)

4725 - 16 Swab Sample
Habitat ID: Date: / 30/ 2021
Species:
Ambystoma mavortium
Stage(s): [] Larval Muvenile or adult
of Individuals in Tank: 🤱
Surface area of tank bottom: 500 cm2
Substrate in bottom of tank is:
[] Bark or fiber [] Rock or gravel [] Soil or dirt
Paper towel [] Other:
Structures in Tank:
[] Live plants [] Artificial plants
[] Wood [] Stones 🕅 Other:

Figure 1. Example label for swab

- 3. Swab the first animal
 - Open the swab
 - Hold the animal firmly so that it will not escape
 - It can be helpful to place the animal in a sandwich bag, as seen in *Figure* 2, to limit its movement. If you decide to do this, use a clean bag for each animal.

- Run the swab along the animal's skin, pushing enough to bend the swab-handle
 - Roll the swab against the direction of movement
- Follow this pattern (Figure 2):
 - 5 times on dorsal surface/back
 - 5 times on ventral surface/belly •
 - 5 times on each foot
 - 5 times on inguinal region/drink patch/around the cloaca
- Place swab in the holder to air dry for a few minutes (Figure 3)



swabbed

- 4. Repeat until all animals are swabbed (up to five individuals)
 - Do not sample the same animal twice, please. Set aside those animals you have • swabbed until you are finished
- 5. Once the last swab is air dried for a few minutes, place all swabs in the labelled WhirlPak bag, swab side down (away from the opening)
- 6. Add three desiccant packs
- 7. Roll up end of bag and twist twist-ties at ends to seal the bag
- 8. Move on to the next tank or container on the list
 - Be sure to change gloves

Sampling water from *aquatic* habitats

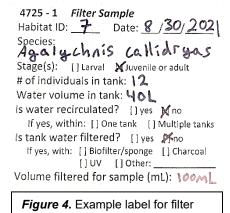
Instructional video here:



https://youtu.be/uMAYfNVY8sA

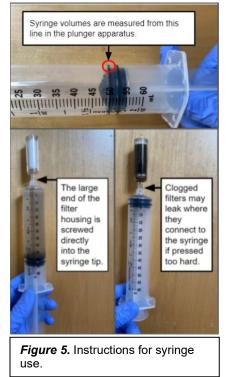
Overview: We want to collect DNA-bearing materials (skin sheds, mucous, crud, etc.) from the water housing animals (eDNA). We want a single eDNA sample per tank. Filtering more water is "better," but please do not break the filter (or your fingers!) trying to push too much through. We will take what we can get. That being said, please tell us how much water was pushed through the filter and the overall volume in the tank.

- 1. Wear new gloves for each habitat
 - If you have sweaty or clammy hands, you may wish to double-glove to make it easier to change the outer pair of gloves between habitats
- 2. Fill out the label on the WhirlPak bag (please use a fine tip permanent marker as pens will smudge) (Figure 1)
 - Habitat ID (to relate results back to a habitat)
 - Species (Latin binomial, please)
 - Life stage(s) (Larva vs. adult)
 - Number of individuals in the tank
 - Approximate volume of water in tank
 - Please provide units! (Metric is preferred, but we can convert)
 - Leave "Volume filtered (mL)" blank until after filtering

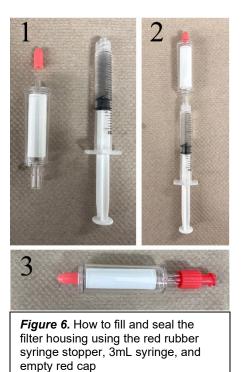


- 3. Open large (60 mL) syringe and filter from wrappers
 - keep the filter wrapper so you have a clean spot to place the filter
- 4. Filter water
 - Draw up 50 mL water (use gradations on syringe)

- Screw filter onto filled syringe (*Figure 5*)
- Push water through filter
 - · Please do not filter water back into container



- 5. Repeat, filtering as much water as possible
 - Unscrew filter and put in a clean place (e.g., the filter's wrapper)
 - Draw up an additional 50 mL water
 - Push water through filter
 - Be careful not to break the filter (*Figure 5*)
 - If water becomes very difficult to press through the filter it is likely clogged. Please stop filtering at this point
 - Write the volume of water pressed through the filter on the last line of the WhirlPak bag
- 6. Dry filter, two times
 - Unscrew syringe
 - Draw up ~50mL of air
 - Reattach syringe to filter
 - Push air through filter to remove excess water
 - Repeat
 - Remove large (60mL) syringe and discard
 - Tap the filter outlet on a clean paper towel to dry it
- 7. Seal filter outlet (Figure 6, #1)
 - Take one of the red rubber syringe stopper, and press it over the filter's outlet



- 8. Add filter preservative (*Figure 6, #2*)
 - Unscrew the luer lock cap from the small (3 mL), pre-filled syringe and secure the syringe to the filter
 - Push the liquid from the small syringe gently into the filter housing with the sealed filter tip facing the ground
 - The liquid should *just* fill the filter case
- 9. Fully seal the filter (*Figure 6, #3*)
 - Remove the now empty 3 mL syringe and dispose of it
 - Acquire one of the red luer lock caps and screw its larger end over the remaining filter opening
- 10. Place filled, sealed, filter in labelled WhirlPak bag
 - Roll up end of bag and twist twist-ties at ends to seal the bag
- 11. Move on to the next tank or container on the list

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